

Bianca Dumontel,<sup>a</sup> Marta Canta,<sup>a</sup> Francesca Susa,<sup>a</sup> Luisa Racca,<sup>a</sup> Andrea Ancona,<sup>a</sup> Nadia Garino,<sup>a</sup> Tania Limongi,<sup>a</sup> Angelica Chiodoni,<sup>b</sup> and Valentina Cauda<sup>a</sup>

<sup>a</sup>Department of Applied Science and Technology, Politecnico di Torino, C.so Duca degli Abruzzi 24, 10129 Turin, Italy  
<sup>b</sup>Center for Sustainable Future Technologies CSFT@Polito, Istituto Italiano di Tecnologia, Corso Trento 21, 10129-Torino, Italy

\*e-mail: [bianca.dumontel@polito.it](mailto:bianca.dumontel@polito.it); Web site: [www.polito.it/TNHlab](http://www.polito.it/TNHlab)

## MOTIVATION

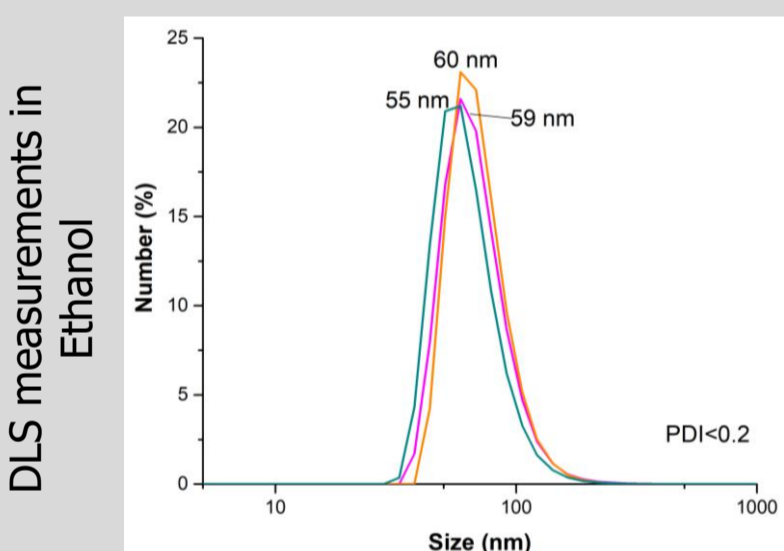
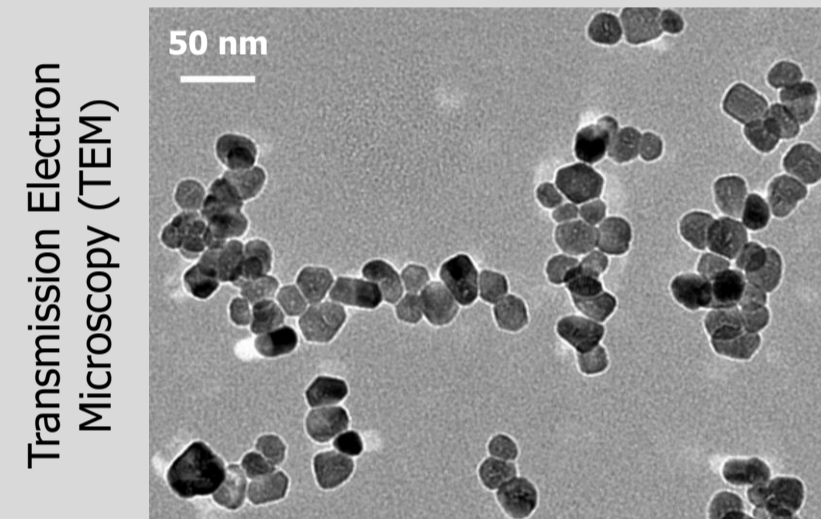
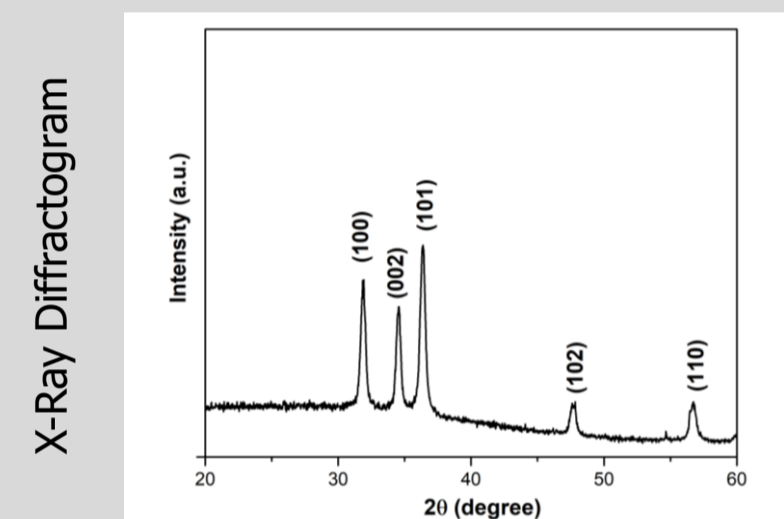
Zinc oxide nanocrystals (ZnO NCs), thanks to their unique properties such as intrinsic cytotoxicity and easy synthesis and functionalization, represent an interesting diagnostic and therapeutic tool for cancer treatment. In order to enable their use in clinical applications, a better control of their chemical and colloidal stability in the biological environment and of the material's biocompatibility is required.

In this study we propose to modify the surface of pristine ZnO NCs with a biomimetic phospholipidic shell, constituted by self-assembled liposomes or cell-derived extracellular vesicles, in order to promote the stability and biocompatibility of the nanocrystals in the physiological environment.

## SYNTHESIS and CHARACTERIZATION

### MICROWAVE-ASSISTED SYNTHESIS

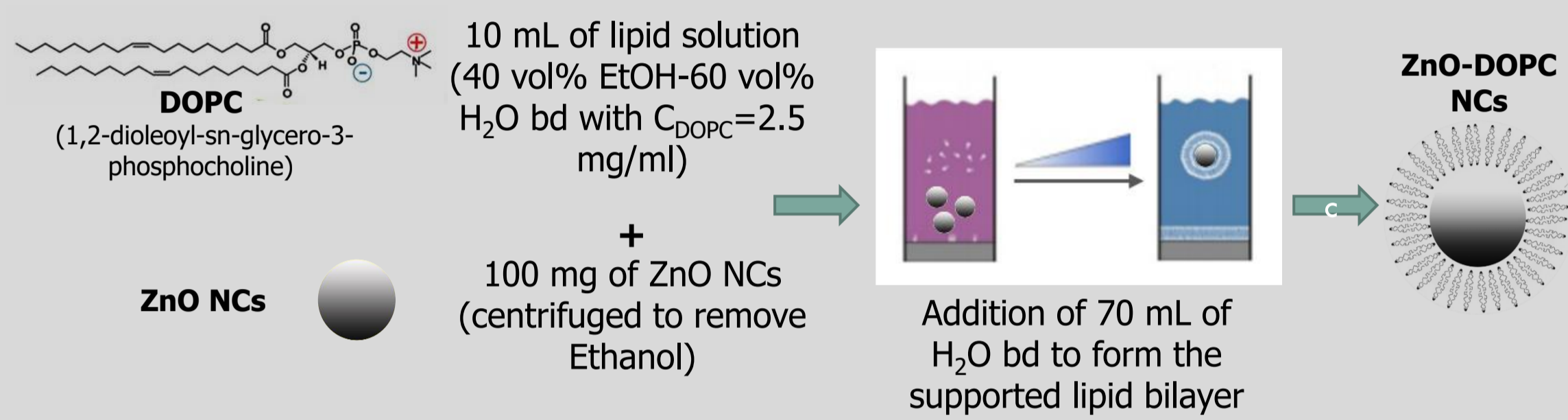
- Sol 1:** 0.1 M Zinc Acetate di-hydrate in Methanol  
 +480  $\mu$ L H<sub>2</sub>O bd (**nucleation promoter**)  
**Sol 2:** 0.2 M KOH in Methanol
- Synthesis mixture in microwaves oven at 60°C for 30 minutes
  - Two washing steps (centrifugation at 3046 rcf for 10 min and redispersion in Ethanol)



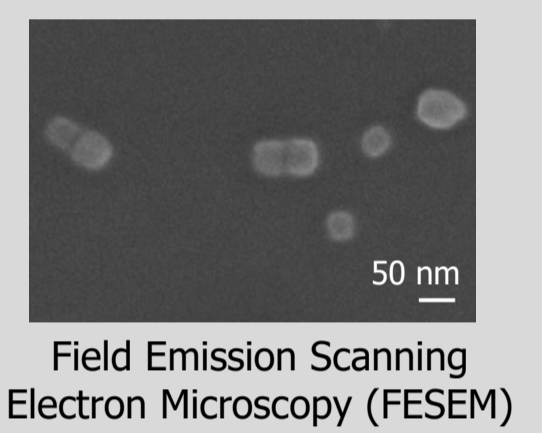
- ZnO NCs**
- Monocrystalline particles of about 20 nm.
  - Homogeneous and monodisperse size distributions.

## LIPID-SHELL FUNCTIONALIZATION with SYNTHETIC LIPIDS

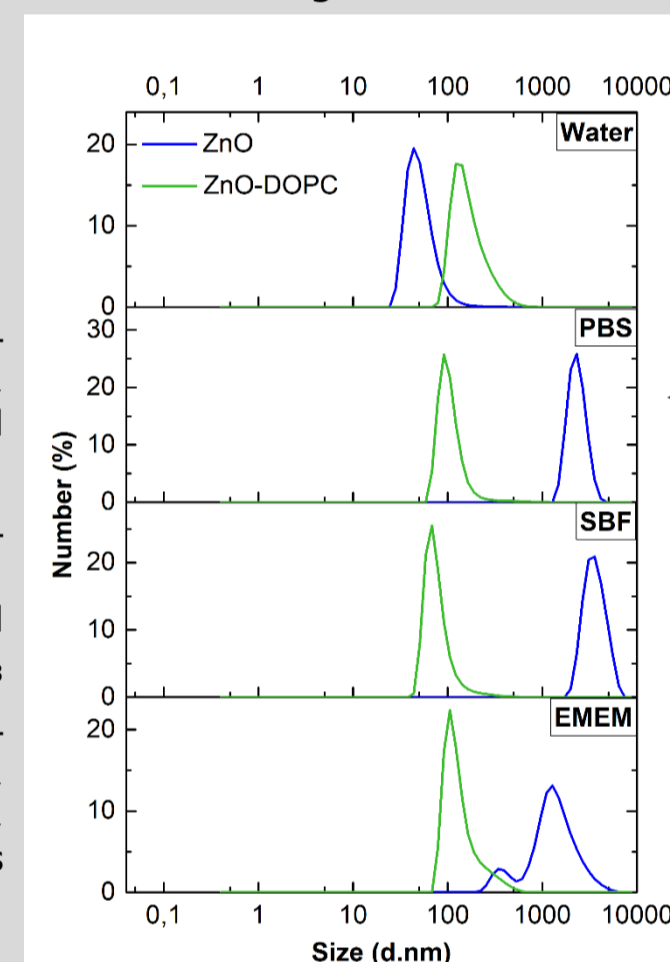
### SOLVENT EXCHANGE METHOD



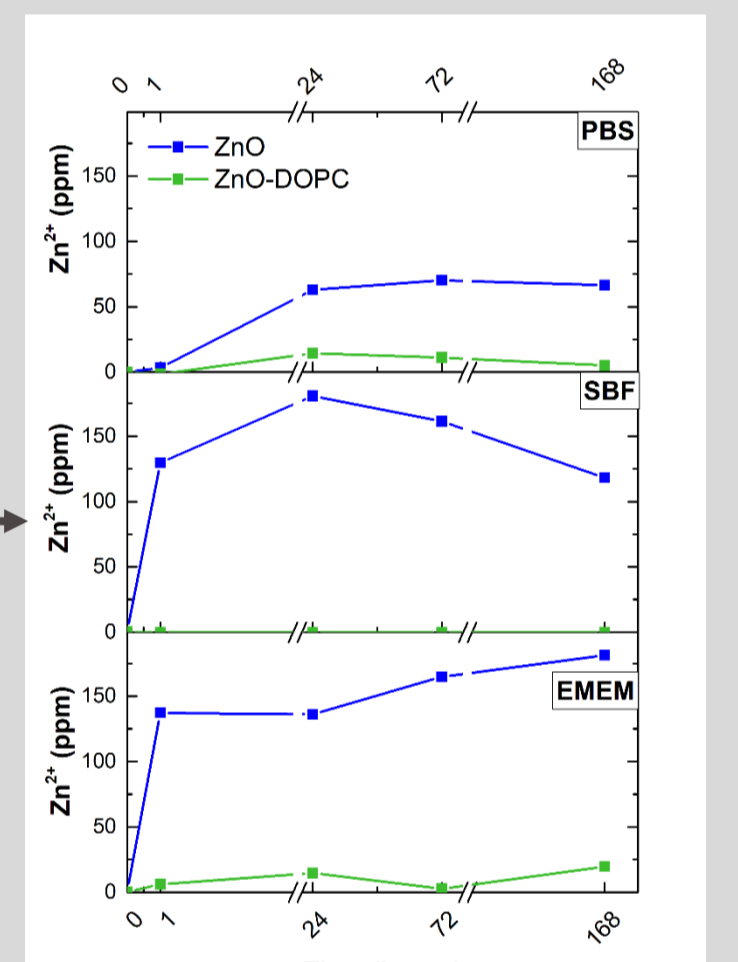
	Z-Pot in water
ZnO NCs	26 mV
DOPC	-15 mV
ZnO-DOPC NCs	1.3 mV



DLS measurements in water and biological fluids



ICP/MS Measurements



Lipid-shell efficiently prevents:  
 NPs aggregation  
 NPs dissolution into Zn<sup>2+</sup>  
 → improves the **BIOSTABILITY**

- Phosphate Buffered Saline (PBS)** – Composition: Na<sub>2</sub>HPO<sub>4</sub>, KH<sub>2</sub>PO<sub>4</sub>, KCl, NaCl
- Simulated Body Fluid (SBF)** – Composition: NaCl, NaHCO<sub>3</sub>, KCl, K<sub>2</sub>HPO<sub>4</sub>, 3H<sub>2</sub>O, MgCl<sub>2</sub>·6H<sub>2</sub>O, CaCl<sub>2</sub>, Na<sub>2</sub>SO<sub>4</sub>, HCl and NH<sub>2</sub>C(CH<sub>2</sub>OH)<sub>3</sub>
- Cell culture medium + 10% FBS** – Composition: CaCl<sub>2</sub>, MgSO<sub>4</sub>, KCl, NaHCO<sub>3</sub>, NaCl, NaH<sub>2</sub>PO<sub>4</sub>, aminoacids, vitamins, glucose, plasma proteins, hormones

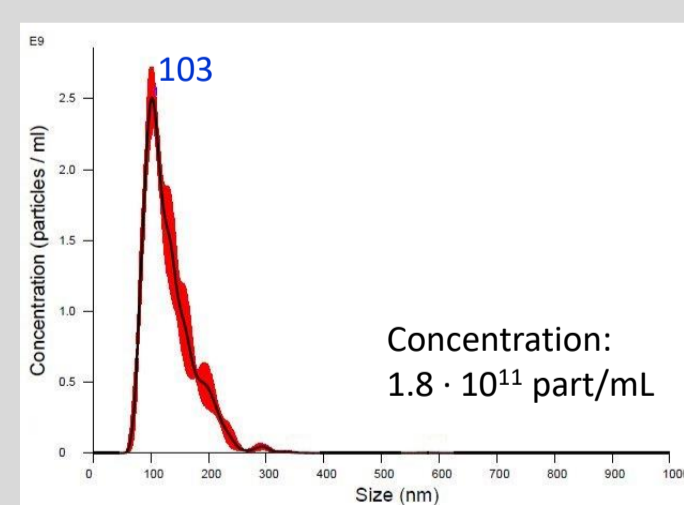
## AUTOLOGOUS EXTRA-CELLULAR VESICLES to improve the BIOCOMPATIBILITY

### EXTRA-CELLULAR VESICLES (EVs)

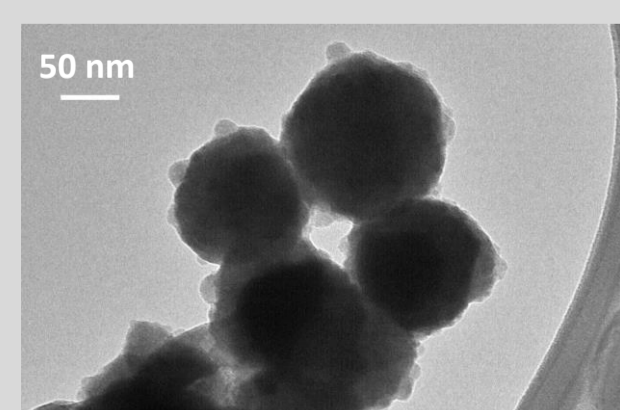
Heterogeneous group of cell-derived membranous structures (comprising exosomes and microvesicles) present in many biological fluids.

- ✓ Autologous -> low immunogenicity
- ✓ Natural stability in blood
- ✓ Biocompatible and non-toxic
- ✓ Small size -> ability to cross biological barriers
- ✓ Intrinsic targeting ability

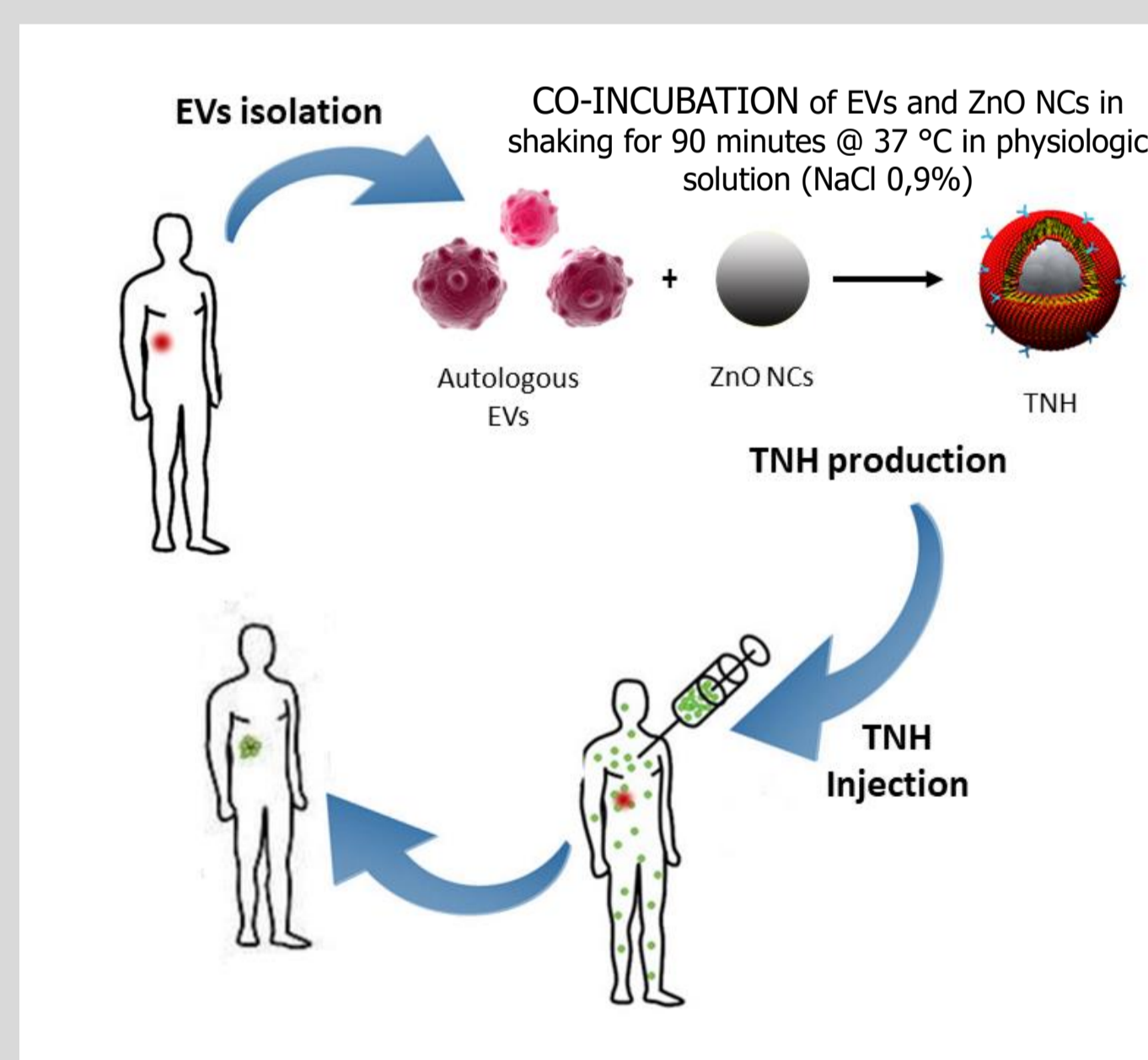
Nanoparticle Tracking Analysis (NTA)



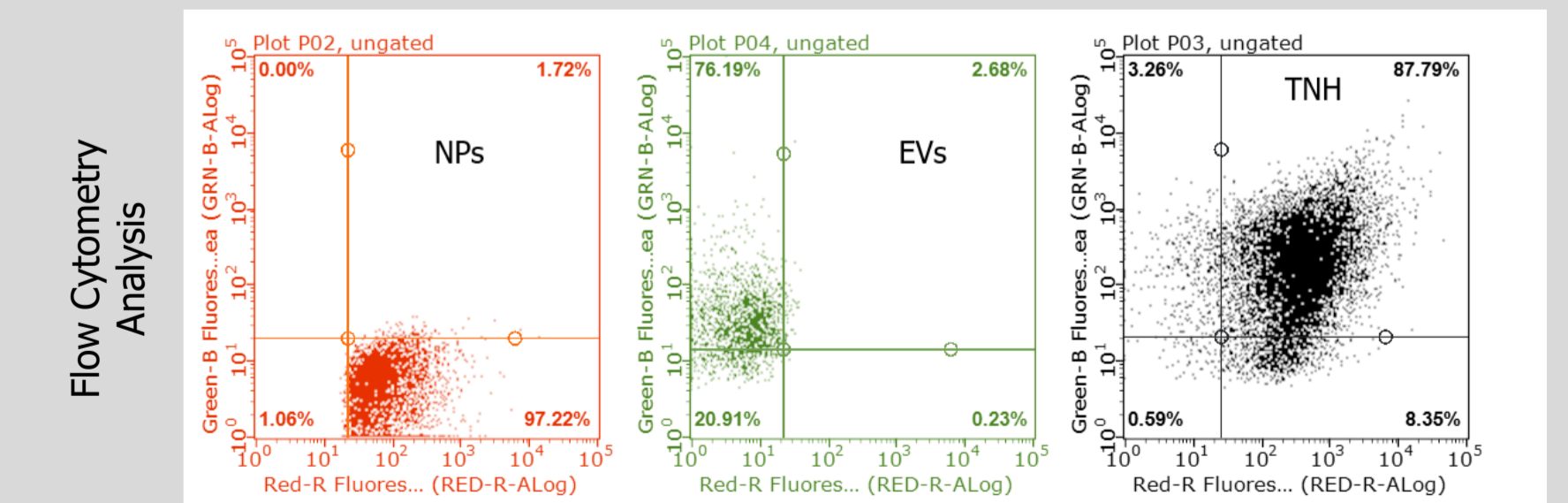
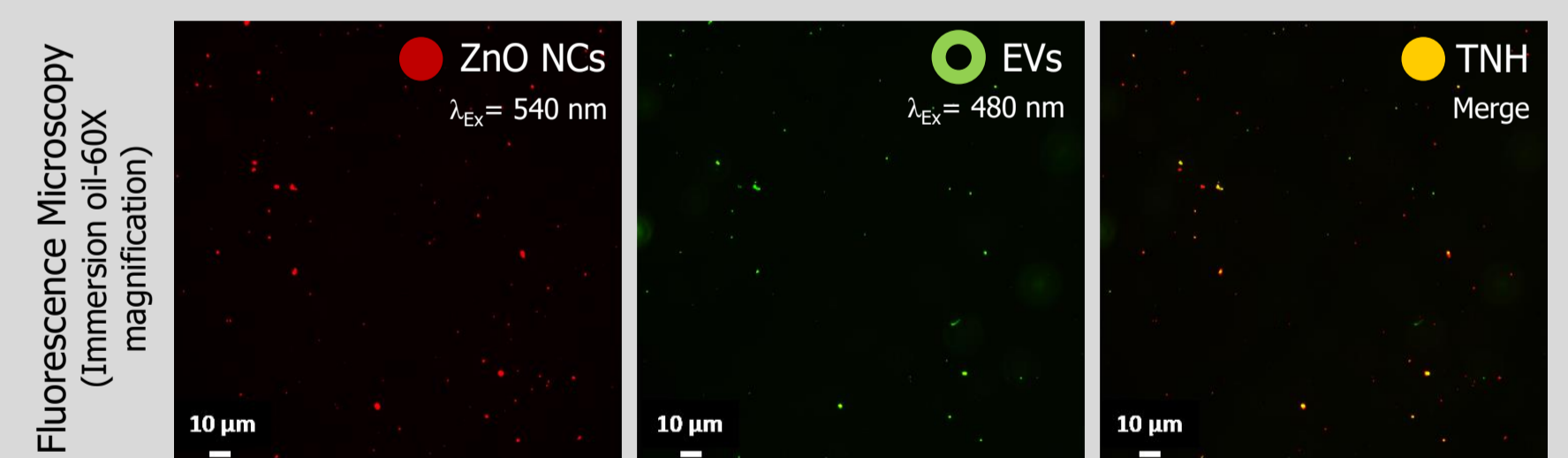
Transmission Electron Microscopy (TEM)



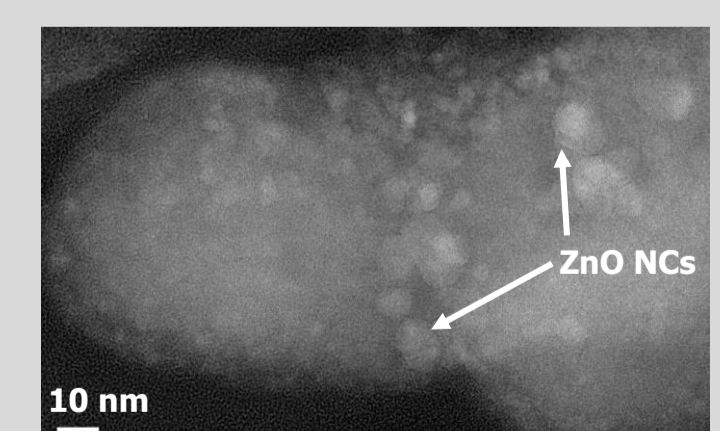
### COUPLING and THERAPEUTIC STRATEGY



### TNH CHARACTERIZATION



Scanning Transmission Electron Microscopy (STEM)



## CONCLUSIONS AND FUTURE OUTLOOK

- Lipid-shell functionalization (DOPC) enables the stabilization of ZnO NCs in biological media and prevents their degradation.
- It is possible to create an hybrid nanoconstruct (TNH) formed by ZnO NCs and cell-derived vesicles (EVs) that, thanks to their biological nature, can provide an enhancement of the biocompatibility.
- Optimisation of TNH production protocol and further characterization of its morphology, bio-stability and bio-compatibility.
- Detailed analysis of therapeutic and imaging capabilities of TNH.

## Bibliography:

- [1] L. Racca, M. Canta, B. Dumontel, A. Ancona, T. Limongi, N. Garino, M. Laurenti, G. Canavese, and V. Cauda, *Smart Nanoparticles for Biomedicine* **2018**, 1, 171-187.
- [2] T. Moore, L. Rodriguez-Lorenzo, V. Hirsch, S. Balog, D. Urban, C. Jud, B. Rother-Rutishauser, M. Lattuada, and A. Petri-Fink, *Chem. Soc. Rev.* **2015**, 44, 6287-6305.
- [3] B. Dumontel, M. Canta, H. Engelke, A. Chiodoni, L. Racca, A. Ancona, T. Limongi, G. Canavese, and V. Cauda, *J. Mater. Chem. B* **2017**, 5, 8799-8813.